

Polymers in Solution

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5. Molar mass determination

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1. Overview





Aggregation (in solution): Dynamic light scattering , field-flow fractionation, small-angle X-ray, fluorencesce spectroscopy, UV-VIS spectroscopy

1. Overview

Molar Mass in Polymers

Number average molar weight

$$\mathbf{M}_{n} = \frac{\sum_{i} c_{i}}{\sum_{i} (c_{i}/Mi)} = \frac{\sum_{i} N_{i}M_{i}}{\sum_{i} N_{i}}$$

Weight average molar weight

$$\mathsf{M}_{\mathsf{w}} = \frac{\sum_{i} (ciM_{i})}{\sum_{i} c_{i}}$$

z-average molecular weight

$$\mathsf{M}_{\mathsf{z}} = \frac{\sum_{i} (ciM_{i}^{2})}{\sum (c_{i}M_{i})} = \frac{\sum_{i} z_{i}M_{i}}{\sum z_{i}}$$

MMD (polydispersity) is with M_w/M_n

For **monodisperse** samples $M_w/M_n = 1$

Polydisperse polymers have $M_w/M_n > 1$





2. Methods for determining the molar mass of macromolecules



Method	Molar mass average	Molar mass range
Absolut method		
Ebulliometry, cryoscopy	M _n	M < 5 x 10 ³
Membrane osmometry	M _n	10 ⁴ < M > 10 ⁶
Isothermal distillation	M _n	M > 5 x 10 ⁴
Sedimentation velocity	$M_{n,}M_{w,}M_{z}$	M> 10 ²
Equilibrium sedimentation	$M_{w,}M_{z}$	M > 10 ²
Vapor pressure osmosis	M _n	M < 2 x 10 ⁴
Static Light Scattering	M _w	M > 5 x 10 ²
Turbidity measurements	M _w	M > 5 x 10 ²
Small-angle X-ray scattering	M _w	M > 5 x 10 ²
Small-angle neutron scattering	M _w	M > 5 x 10 ²
Dynamic Light Scattering	M _w	M > 5 x 10 ²
Mass spectroscopy- MALDI-TOF	$M_{n,}M_{w,}M_{z}$	M > 5 x 10
Equivalent method		
End-group analysis- (titration, NMR, IR)	M _n	M < 5 x 10 ⁴
Relative method		
Dilute solution viscometry	Μ _η	M > 10 ²
Gel Permeation chromatography	$M_{n,}M_{w,}M_{z}$	M < 10 ⁷
Supercritical fluid chromatography	$M_{n,}M_{w,}M_{z}$	M < 10 ⁷
Field-flow fractionation	M_{n} , M_{w} , M_{z}	M > 10 ³

2. Methods for determining the molar mass of macromolecules



Characterizing polymer structure with small-angle neutron scattering: A Tutorial



FIG. 5. Illustration of polymers with (left to right) globular, ideal, and swollen conformations. The corresponding Porod plots and slopes are shown below each.

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3. Colligative properties and M_n

Colligative properties of solutions are properties that depend upon the concentration of solute molecules or ions, but not upon the identity of the solute. **Colligative properties include vapor pressure lowering, boiling point elevation, freezing point depression, and osmotic pressure.**

Cryoscopy

$$\left(\frac{\Delta T_f}{C}\right)_{C=0} = \frac{RT^2}{\rho \Delta H_f \overline{M}_n} + A_2 C$$

- ΔT_{f} : freezing-point depression,
- C: the concentration in grams per cubic centimeter
- R: gas constant
- T: freezing point
- ΔH_{f} : the latent heats of fusion
- A2 : second virial coefficient

 M_n for low molar mass ($\leq 10000 \text{ g/mol}$)

 $\left(\frac{\Delta T_b}{C}\right)_{C=0} = \frac{RT^2}{\rho \Delta H_v \overline{M}_n} + A_2 C$

Ebulliometry

 ΔT_b : boiling point elevation ΔH_v : the latent heats of vaporization



5. Molar mass determination

3. Colligative properties and M_n

Membrane osmometry METHODS BASED ON COLLIGATIVE PROPERTIES

SIMPLE MEMBRANE OSMOMETER



$$10^4 < M_n > 10^6$$

$$\frac{\pi}{c_B} = RT\left(\frac{1}{M_n} + A_2 c_B\right)$$

Determination of M_n (interception) and A₂ (slope)



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3. Colligative properties and M_n

Vapour pressure osmometry METHODS BASED ON COLLIGATIVE PROPERTIES



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M_n for low molar mass ($\leq 20000 \text{ g/mol}$)

4. Equivalent method

End group analysis

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$M_n \leq 50000 \text{ g/mol}$

- Molecular weight limitation up to 50000 g/mol (the concentration of end groups has to be sufficient to get an accurate measurement)
- You have to know how many end groups there are per molecule (to find molar mass), OR you know the molar mass, and want to know number of end groups per molecule. Limitation for branched polymers
- End-group must have detectable species
- a. Vinyl polymer: -CH=CH₂
- b. Ester Polymer: -COOH, -OH
- c. Amide and urethane polymer: -NH₂, -NCO
- d. Radioactive isotopes or UV, IR, NMR detectable functional groups



4. Equivalent method



How to Determine Hyaluronic Acid Molecular Weight Using Gel Electrophoresis

Hyaluronic Acid, is a natural non-sulfated glycosaminoglycan produced in many organs and tissues. It was discovered in the 1930s and was originally thought to have no physiological function except serving as a lubrication "space filler" between joints. With additional research, hyaluronic acid is now appreciated as an important part of the extracellular matrix. Indeed, it has critical roles in many cell signaling events such as proliferation, inflammation, wound healing, and fertilization.



Figure 1. Repeating disaccharide units of Hyaluronic Acid



Figure 2. Source of Agarose

Select-HA[™] Ladders were run on 1% agarose gel prepared using 2 different agarose source. Both agarose gels were run using same conditions.

Agarose gel prepared using Source #1 resulted with more defined HA bands than agarose gel prepared using Source #2.

Lane 1 – 5 µL Select-HA™ HiLadder Lane 2 – 5 µL Select-HA™ LoLadder

4. Equivalent method





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Li Zihao, Jiang Yanyan, Wüst Kilian, Callari Manuela, Stenzel Martina H. (2020) Crosslinking of Self-Assembled Protein-Polymer Conjugates with Divanillin. Australian Journal of Chemistry 73, 1034-1041.



The intensity of scattered light is directly proportional to the molar mass.

The angular dependency of scattered light is proportional to the size (radius).



Static light scattering From Rayleigh Ratio to molar mass

Zimm Equation, J. Chem. Phys. 16, 1093-1099 (1948).

$$\frac{K^*c}{R(\theta)} = \frac{1}{M_w}P(\theta) + 2A_2c$$

$$K^* = 4\pi^2 (dn/dc)^2 n_0^2 N_A^{-1} \lambda_0^{-4}$$



The amount of scattered light at scattering angle 0 is directly proportional to the product of **molar mass** (g/mol) and concentration (g/ml).

MALDI-TOF

The Matrix Assisted Laser Desoption Ionization Time of Flight Mass Spectroscopy (MALDI-TOF-MS) is a fast and sensitive absolute method for determining both the num-ber average and weight average molar masses. It has a special status in the analysis of polymers of biological origin. In an ideal case, molar masses of <300,000 g/mol can be measured

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Fig. 3.16 MALDI-TOF-MS-spectrum of a polystyrene. The relative abundance (normalized to the most abundant peak) is plotted against the mass/charge ratio (m/z). The numbers above the peaks are the relative abundance and the corresponding m/z value

Ultracentrifuge

The ultracentrifuge (UC) is a centrifuge which rotates at very high speeds and was origi-nally developed by Svedberg for his research on inorganic and organic colloids (Svedberg and Pedersen 1940). Three types of experiment are possible, which allow conclusions about **the shape, conformational**

changes, and size distribution of dispersed particles or dissolved macromolecules.

- (1) Analysis of Sedimentation Velocity
- (2) Measurement at Thermodynamic Equilibrium
- (3) Sedimentation Equilibrium in a Density Gradient





- 5. Molar mass determination
 - 6. Relative method

Dilute solution viscometry

A. IUPAC suggested the terminology of solution viscosities as following. Relative viscosity: ηt

$$\eta_{rel} = \frac{\eta}{\eta_o} = \frac{t}{t_o}$$

$$\eta_o: \text{ solution viscosity}$$

$$\eta_o: \text{ solvent viscosity}$$

$$t: \text{ flow time of solution}$$

$$t_o: \text{ flow time of solvent}$$

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Specific viscosity:

$$\eta_{sp} = \frac{\eta - \eta_o}{\eta_o} = \frac{t - t_o}{t_o} = \eta_{rel} - 1$$

Reduced viscosity:

$$\eta_{rel} = \frac{\eta_{sp}}{c} = \frac{\eta_{rel} - 1}{c}$$

Inherent viscosity: $_{C}$ $\eta_{inh} = -\frac{1}{2}$

$$\eta_{inh} = \frac{\ln \eta_{rel}}{c}$$

Intrinsic viscosity:
$$[\eta] = (\frac{\eta_{sp}}{c})_{c=o} = (\eta_{int})C = 0$$

Dilute solution viscometry



$$\left(\frac{M^2}{[\eta]}\right)^{1/3} = A_\eta + B_\eta M^{1/2}$$

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Figure 5 Sketch of the (a) Oswald viscometer, (b) Ubbelohde dilution viscometer in the normal form, and (c) compact form.

$[\eta] = KM^a$ Mark–Houwink– Sakurada equation

- *K*, *a* = constants of Mark-Houwink equation for given polymer, solvent and temperature; *M* = molar mass
- Traditional method of the determination of molar mass
- Viscosity average (M_v) close to weight-average (M_w)





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Molar Mass from Mark-Houwink Equation



Molar mass versus elution volume plots of linear polystyrene by MALS and calculated from Mark-Houwink equation.

Dilute solution viscometry

Exponent of Mark-Houwink equation

- \rightarrow Linear macromolecules in thermodynamically good solvents: $a \approx 0.7$
- \rightarrow Linear macromolecules in thermodynamically poor solvents: $a \approx 0.5$
- → Oligomers: $a \approx 0.5$
- → Hard spheres: a ≈ 0
- → Extended chains: $a \approx 0.8$ to ≈ 1.5
- → Linear polymers have linear MH plots
- → Curved plots indicates branching



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Dilute solution viscometry



Mark-Houwink plots of epoxy resin, linear polystyrene, linear poly(methyl methacrylate, linear poly(benzyl methacrylate), linear poly(iBuPOSSMA) and star-branched poly(isobutyl methacrylate).

Dilute solution viscometry

How Does the Branching Effect of Macromonomer Influence the Polymerization, Structural Features, and Solution Properties of Long-Subchain Hyperbranched Polymers?





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Types of liquid chromatographic separations



Size Exclusion Chromatography (SEC)

Liquid Chromatography under Critical Conditions (LCCC) Liquid Adsorption Chromatography (LAC)

Types of liquid chromatographic separations

Liquid Chromatography under Critical Conditions (LCCC)



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- Large particles cannot enther gel and are excluded. They have less volume to traverse and elute sooner
 - Small particles can enter gel and have more volume to traverse. They elute later



Size Exclusion Chromatography (SEC)



SEC is separating polymers by their **hydrodynamic volume** or **hydrodynamic radius** – which is affected by various things, in particular the polymer (chemistry and structure), solvent, solvent/polymer interactions, and temperature.



Compact, more tightly coiled, smaller hydrodynamic volume



More elongated, larger hydrodynamic volume



Size Exclusion Chromatography (SEC)- Calibration curve



What is SEC-MALS?

• **SEC** = **S**ize-**E**xclusion **C**hromatography

Size-Exclusion Chromatography (SEC) is a chromatographic method in which molecules are separated based on their size, or, in more technical terms, their hydrodynamic volume.

• MALS = Multi-Angle Light Scattering

Two modes of operation

$$I_s(\theta) \propto c \times M \times \left(\frac{dn}{dc}\right)^2$$

- A batch experiment measures an unfractionated sample.
 - \rightarrow Only the weight-averaged molar mass M_w, z-averaged RMS radius R_z, and potentially A₂ will be determined.
 - \rightarrow No information about the polydispersity of the sample can be obtained.
- SEC allows the sample to be chromatographically separated.
 - \rightarrow Molar Mass and RMS Radius moments and distributions can be assessed.

SEC-MALS-UV/RI method

 SEC provides separation and the molar mass is measured by online MALS and concentration detectors. Molar mass moments (M_n, M_w, M_z), radius moments, dispersity, conformation

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$$I_{s}(\theta) \propto c \times M \times \left(\frac{an}{dc}\right)^{-1}$$



BSA

Dextran







molar mass vs. time

SEC-MALS-UV/RI method

- Knowledge of *dn/dc* is necessary for
 - → Molar mass determination from light scattering measurements
 - → Determining concentrations by an RI detector
 - → Analog RI calibration
- The value of *dn/dc* depends on
 - → Polymer composition
 - → Solvent
 - → Molar mass
 - → Laser wavelength
- The *dn/dc* can be obtained from
 - → The literature
 - → Direct measurement with an RI detector (online or offline)
- For greatest accuracy, the dn/dc used in light scattering experiments should have been obtained for your polymer, in your solvent, and at the wavelength of your LS detector.

- *dn/dc* describes the change in the refractive index of a solution as a function of solute concentration.
- dn/dc has units of mL/g; it expresses how much the refractive index of a solution theoretically increases for every g of solute contained in a 1 ml final volume of solution.
- The *dn/dc* of a polymer solution should be measured at polymer concentrations appropriate for chromatography conditions; some non-linearity can occur at very high solute concentrations.
- To determine molar mass by LS, dn/dc needs to be known
 - → For your solute
 - → In your solvent
 - \rightarrow At the wavelength of your LS detector
 - \rightarrow Even when you are not using a concentration detector!



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Typical SEC-MALS-hardware setup



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Field-flow fractionation (FFF)



It is a separation technique in which a field (thermal, electric, magnetic, hydraulic, gravitational, ...) is applied to a diluted <u>suspension</u> in a fluid or to a <u>solution</u> pumped through a long and narrow channel, perpendicular to the direction of the field, in order to cause the separation of particles present in the fluid, depending on their differing "mobilities" under the force exerted by the field. The FFF method is unique to other separation techniques because it can separate materials over a wide <u>colloidal</u> size range while maintaining high resolution. Although FFF is an extremely versatile technique, there is no "one size fits all" method for all applications.

5. Molar mass determination

7. Polymer bioconjugates: Modern design concepts toward precision hybrid materials





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https://doi.org/10.1016/j.progpolymsci.2020.101241

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